Microfluidics Assisted Fabrication of Three-Tier Hierarchical Microparticles for Constructing Bioinspired Surfaces

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Supporting Information

ABSTRACT: Construction of textured bioinspired surfaces with refined structures that exhibit superior wetting properties is of great importance for many applications ranging from self-cleaning, antibiofouling, anti-icing, oil/water separation, smart membrane, and microfluidic devices. Previously, the preparation of artificial surfaces generally relies on the combination of different approaches together, which is a lack of flexibility to control over the individual architecture unit, the surface topology, as well as the complex procedure needed. In this work, we report a method for rapid fabrication of three-tier hierarchical microunits (structures consisting of multiple levels) using a facile droplet microfluidics approach. These units include the first-tier microspheres consisting of the second-tier close-packed polystyrene (PS) nanoparticles decorated with the third-tier elegant polymer nanowrinkles. These nanowrinkles on the PS nanoparticles are formed according to the interfacial instability induced by gradient photopolymerization of N-isopropylacrylamide (NIPAM) monomers. The formation process and topologies of nanowrinkles can be regulated by the photopolymerization process and the fraction of carboxylic groups on the PS nanoparticle surface. Such a hierarchical microsphere mimics individual units of bioinspired surfaces. Therefore, the surfaces from self-assembly of these fabricated two-tier and three-tier hierarchical microunits collectively exhibit “gecko” and “rose petal” wetting states, with the micro- and nanoscale structures amplifying the initial hydrophobicity but still being highly adhesive to water. This approach offers promising advantages of high-yield fabrication, precise control over the size and composition of the microspheres, and integration of microfluidic droplet generation, colloidal nanoparticle self-assembly, and interfacial polymerization-induced nanowrinkles in a straightforward manner.

KEYWORDS: three-tier, hierarchical structure, microfluidics, nanowrinkle, bioinspired surface

Wrinkled surfaces in nature have inspired scientists to mimic them with artificial topological surfaces with high spatial periodicity. Such structures on the micro- and nanoscale show optical, biological, electronic, or acoustic properties due to their wave interference phenomena, exceptional hydrodynamic properties, enhanced adhesion force, low surface energy, and/or large specific surface area.1,2 The integration of wrinkled surfaces with multiple hierarchical (i.e., structured on multiple scales in one unit) architectures even further improves the performance in catalytic support, electrodes, controllable superwetting modulation, digital microfluidics, microfluidic sieves, surface enhanced Raman spectroscopy (SERS), tracking and identification technology, flexible electronics, diffraction gratings, separation science, drug delivery, tissue engineering, industrial field-like abrasives material, and environment protection materials.3 Therefore, study and construction of the unit of such materials/surfaces become prominent to understand the relationship between structures and functionalities.
A summary of various methods for fabrication of artificial hierarchical structures on two-dimensional (2D) polymer films with wrinkled surfaces has been reported by Chen and Yang. The underlying mechanism of the wrinkle formation on the film is attributed to the applied stress on their surface. When this stress is larger than a critical value, it leads to an interfacial instability due to the mismatch of mechanical properties like the Young’s modulus in the equilibrium state between different layers—normally between a stiff skin layer and soft substrate. The applied stress is commonly raised by mechanical (e.g., stretching, compressive force), thermal (e.g., electron irradiation), physical (e.g., osmotically driven), and chemical (e.g., solvent swelling, ultraviolet (UV) irradiation, oxygen plasma treatment) approaches.

Recently, hierarchical architectures have been developed for three-dimensional (3D) materials and have attracted intense attention. The functional properties (defined as stimuli-responsive) of 3D hierarchical structures are determined by their chemical composition and tailored surface topology, exhibiting the advantages of high spatial frequency, enlarged specific surface roughness and area. In particular, controlling of surface topology enables the control of interfacial phenomena such as fluid wetting, flow, and adhesion. These hierarchical structures have enhanced functionalities by their surface topology on the micro- and nanoscopic length scale, thus broadening their applications in many fields. For example, regarding the application of wetting materials, tailoring surface roughness can give rise to an amplified hydrophilicity/hydrophobicity for the film surfaces. Accordingly, an effective approach for fabricating superwetting surfaces is to create hierarchical structures. Hierarchical bioinspired structures that are, for example, the mimicry of gecko feet, a mosquito eye, and a rose petal have been fabricated through either the top-down methods of lithography and laser writing or through bottom-up approaches like self-assembly of colloidal particles.

Artificially wrinkled surfaces on particles are typically constructed in two separate steps, that is, synthesis of the polymer template and postprocessing. The polymer templates can be synthesized using flow lithography, electro-spraying, or microfluidic techniques. Subsequently, the exposed surfaces are modified by using additional material coating, plasma etching, surface chemical oxidation, chemical modification, solvent evaporation, electron irradiation, or UV irradiation. Although effective, these fabrication methods are time-consuming and require complex processes. It is worth mentioning that among these technologies, the droplet-based microfluidic technique stands out by its production of droplet templates with high uniformity in size and tunability in particle geometry and chemical composition. Moreover, this method can be easily integrated with other approaches (as a hybrid approach: combination of top-down and bottom-up method), thus enabling continuous construction of 3D hierarchical microparticles with different chemical and physical properties. Liu et al. reported a droplet-based microfluidic method for fabricating polymer micro-
particles. The surface textures on polymer microparticles could be tuned via interfacial instability triggered by emulsion droplets shrinking. Metal nanoparticle-loaded polymer composite particles have also been synthesized via microfluidic approach integrated with in situ photopolymerization and photoreduction. These composite particles could serve as sensing units for molecular detection. However, these microparticles only show two-tier hierarchy of nanostructures on the microparticles.

In this work, we report a hybrid approach for rapid construction of three-tier hierarchical microspheres, using a droplet-based microfluidic technique to produce microparticles with patterned nanoparticles (via colloid nanoparticle self-assembly) covered with a convoluted (wrinkled) surface (via gradient photopolymerization). The fabricated three-tier microspheres can be applied as uniform units to form bioinspired surfaces. However, these microparticles only show two-tier hierarchy of nanostructures on the microparticles.

RESULTS AND DISCUSSION

Fabrication of Three-Tier Hierarchical Units of Microspheres Consisting of Nanoparticles Covered with Nanowrinkles. Three-tier hierarchical microspheres with nanowrinkles were successfully fabricated using droplet microfluidics. A microfluidic device with a flow-focusing geometry was used to generate water-in-oil emulsion droplets with high uniformity (coefficient of variation of 1.5%) and high yield (>1000 microspheres per second). Herein, a suspension containing 3 w/v% carboxylic-functionalized polystyrene (f-PS) nanoparticles, 5 wt % NIPAM as curable monomer, and 0.5 wt % N,N-methylene bis(acrylamide) (MBA) as cross-linker was used as the water phase (droplet). The oil (continuous) phase was n-hexadecane with 19 wt % sorbitane monooleate sorbitan oleate (Span80) as surfactant to stabilize the produced droplets and 5 wt % 2,2-diethoxyacetophenone (DEAP) as a photoinitiator to initiate the photopolymerization reaction. The concentrations of NIPAM and MBA in the water phase and Span80 and photoinitiator in the oil phase were kept constant in all experiments. The water phase was sheared off to form monodispersed emulsion droplets with the oil phase at the flow-focusing area. Generated droplets were collected in a beaker containing oil phase, as shown in Figure 1a.

Figure 2. (a) AFM image of the surface of a f-PS@MS and (b) the corresponding dimensions measured along the red dashed line. (c) HR-SEM (scale bar: 1 μm) and a close-up image (scale bar: 200 nm) of the cross-sectional morphology of a fabricated f-PS@MS cut by using a focus ion beam.
assembled, driven by interfacial tension and electrostatic repulsion,\textsuperscript{51} producing close-packed nanopatterns on the droplet surface (Figure 1b). To obtain the well-ordered nanopatterns, the collected droplets in the beaker containing the oil phase were kept for more than 1 h. Afterward, these droplets were photopolymerized through UV-induced interfacial free-radical polymerization (IFRP), yielding solid microspheres (Figure 1c). The volume shrinkage of produced liquid microdroplets into solid microspheres was approximately 96.38% (see the calculation in Supporting Information).

These solid microspheres show a three-tier hierarchical structure, including the first tier of microframe, the second tier of close-packed nanopatterns assembled from f-PS nanoparticles, and the third tier of nanowrinkles on every individual f-PS nanoparticle surface caused by the gradient-UV photopolymerization. Figure 1d shows an optical microscope (OM) image of droplet generation in the microfluidic device and the collected monodispersed droplets observed under the bright field (Figure 1e) and fluorescent field (Figure 1f) modes. The fluorescent image shows bright red color under 365 nm excitation due to the imbedded quantum dots (CdSe/ZnS) in the center of purchased f-PS nanoparticles. High-resolution scanning electron microscope (HR-SEM) images show the three-tier structures of the polymerized microspheres (Figure 1g).

Figure 2 shows the surface of a three-tier hierarchical microsphere (f-PS@MS) and the corresponding dimensions of the nanowrinkles measured by an atomic force microscope (AFM). From the observation, the nanowrinkles are in the sub-30 nm range, and their distribution on every individual nanoparticle is relatively uniform. To further view the inside of the microspheres, we cut a f-PS@MS using a focused ion beam. Figure 2c shows the cross section of the cut surface. It reveals that the microsphere was fully filled with f-PS nanoparticles and nanowrinkles exist on each f-PS nanoparticle. The flat surface of the cut area in the HR-SEM image, hereby, was caused by heat melting during the focus ion beam cutting.

The produced f-PS@MS exhibits a large surface-to-volume ratio of $\sim 3 \times 10^7 \text{m}^{-1}$ (about 47 times larger than that of a smooth microsphere at the same diameter), high spatial frequency of $\sim 3.3 \times 10^5 \text{mm}^{-1}$ (which is defined as the frequency of the nanowrinkle structure repetition per unit of distance), and high-density nanogaps of $\sim 422$ nanogaps per $\mu \text{m}^2$ (a gap between two adjacent peaks of the nanowrinkles is defined as a nanogap) (Figures S1–S2). The surface-to-volume ratio was calculated by multiplying the measured surface area of individual nanoparticles by the number of particles inside a microsphere. The surface area of a nanoparticle with nanowrinkles was calculated based on the AFM measured average nanowrinkle dimensions and gaps. It is therefore worth mentioning that these values might be slightly overestimated. Therefore, these f-PS@MS are expected to have high potential for coating materials (surface wettability control via micro- and nanoscale structures), molecular detection (localized surface plasmon resonance due to the presence of nanogaps), bioassays (high adhesive surface), and catalysis support (large specific surface area).
termed as f-PS@MS. The morphology of these f-PS@MS(T) is shown in Figure 3a. As can be seen, all f-PS nanoparticles assembled into micrometer clusters without wrinkle formation on their surface, compared with that in Figure 1g. The surface morphology of single f-PS nanoparticle was found to be similar to the virgin one (Figure S 3f). This indicates that the volume shrinkage during the water evaporation process from liquid droplets to solid microspheres did not induce wrinkle formation on the f-PS nanoparticle surface.

Moreover, a pure suspension of bare-PS nanoparticles (bare means without carboxylic acid modification) was used as the water phase to generate the droplets, and solid microspheres were subsequently produced via solvent evaporation at 60 °C overnight. These particles were termed as f-PS@MS(T). The morphology of these f-PS@MS(T) is shown in Figure 3a. As can be seen, all f-PS nanoparticles assembled into micrometer clusters without wrinkle formation on their surface, compared with that in Figure 1g. The surface morphology of single f-PS nanoparticle was found to be similar to the virgin one (Figure S 3f). This indicates that the volume shrinkage during the water evaporation process from liquid droplets to solid microspheres did not induce wrinkle formation on the f-PS nanoparticle surface.

Mechanism of Nanowrinkle Formation. As presented in previous literatures, the formation of nanowrinkles is commonly due to the volume shrinkage resulting from solvent evaporation and/or polymerization reactions occurring on the structure surface. To understand the mechanism of nanowrinkle formation in this work, a series of comparative experiments were designed and conducted.

In the first comparative experiment, a suspension of pure f-PS nanoparticles was used as the water phase to produce the droplets, and solid microspheres were subsequently produced via solvent evaporation at 60 °C overnight. These particles were termed as f-PS@MS(T). The morphology of these f-PS@MS(T) is shown in Figure 3a. As can be seen, all f-PS nanoparticles assembled into micrometer clusters without wrinkle formation on their surface, compared with that in Figure 1g. The surface morphology of single f-PS nanoparticle was found to be similar to the virgin one (Figure S 3f). This indicates that the volume shrinkage during the water evaporation process from liquid droplets to solid microspheres did not induce wrinkle formation on the f-PS nanoparticle surface.

Furthermore, to determine the occurrence of chemical reactions between the nanoparticles and the NIPAM monomers, a mixture of bare-PS nanoparticles with the same concentration of NIPAM and MBA was used as the water phase to generate the droplets, which were subsequently photopolymerized by UV irradiation to form microspheres termed as bare-PS@MS. Figure 3c shows the top-view HR-SEM images of a fabricated bare-PS@MS, exhibiting a smooth surface on each bare-PS nanoparticle. It was found that a NIPAM polymer layer filled in the space between the bare-PS nanoparticles (see red arrows in Figure 3c), indicating the occurrence of NIPAM cross-linking reaction. In contrast to f-PS@MS (Figure 1g), the bare-PS@MS (Figure 3c) shows a smooth surface, suggesting a critical role of the surface carboxylic groups on the surface of the PS nanoparticles on the formation of nanowrinkles during photopolymerization-induced solidification from droplets to particles.

To further investigate the effect of the surface carboxylic groups on the photopolymerization process, copolymer polystyrene nanoparticles containing different molar fractions of carboxylic groups were employed to fabricate the microspheres. In four separate experiments, bare-PS (no carboxylic groups), PS-co-PAA1 (containing 10% carboxylic groups in molar fraction), PS-co-PAA2 (containing 18.5% carboxylic groups in molar fraction), and f-PS nanoparticles (fully functionalized with carboxylic groups on the surface) were mixed with 5 wt % NIPAM and 0.5 wt % MBA, as the aqueous phases to prepare hierarchical microspheres denoted as bare-PS@MS, PS-co-PAA@MS, PS-co-PAAh@MS, and f-PS@MS, respectively. The collected microdroplets were suspended in the continuous phase of n-hexadecane containing 19 wt % Span80 and 5 wt % DEAP, followed by polymerization using UV irradiation. Figure 3c–f shows the HR-SEM images of these fabricated bare-PS@MS, PS-co-PAA@MS, PS-co-PAAh@MS, and f-PS@MS. Without carboxylic groups on surface, the bare-PS nanoparticles could assemble to nice and smooth microspheres via photopolymerization (Figure 3c). There was no obvious difference found on both the microspheres and the nanoparticle surfaces. On the surface of PS-co-PAAh@MS in Figure 3d (also Figure S4a), a layer of NIPAM polymer was observed with imprinted traces of the encapsulated nanoparticles. Under this NIPAM polymer layer, PS nanoparticles were closely packed with rough surfaces, indicating that the

Figure 4. Schematic diagram of the formation of wrinkled surfaces on a f-PS@MS. (a) Droplet with close-packed f-PS nanoparticles before curing. (b) Initial state of droplet curing and formation of a thin stiff layer at the top surface. (c) Further gradient-curing (cross-linking reaction) and stress development. (d) Wrinkle formation on the f-PS nanoparticle surface.
cross-linked NIPAM monomers remained attached to the surface of PS-co-PAA@MS. Moreover, on the PS-co-PAA@MS surface in Figure 3e (also Figure S4b), close-packed and well-ordered nanoparticles were observed with obvious nanowrinkled surface. On the f-PS@MS surface, nanowrinkles were found fully covering the nanoparticle and microsphere surfaces, as shown in Figures 3f and 2c. These results suggest that, during the photopolymerization process, the microsphere and nanoparticle surfaces change obviously with the fraction of carboxylic groups of the PS nanoparticles.

By comparing these microspheres constructed by evaporation-induced solidification composed of f-PS (Figure 3a) and bare-PS (Figure 3b) nanoparticles and photopolymerization-induced solidification composed of bare-PS (Figure 3c), PS-co-PAA (Figure 3d), PS-co-PAA@ (Figure 3e), and f-PS (Figure 3f) nanoparticles, we can conclude that both the photopolymerization process and the fraction of carboxylic groups on PS nanoparticle surface play critical roles for the nanowrinkle formation.

Therefore, we propose the following mechanism to explain the wrinkle formation. Chemically, the NIPAM molecule contains an amine group. When there is no carboxylic group on the NIPAM monomers and the bare-PS nanoparticles, as the NIPAM monomers and produced oligomers bind to the nanoparticle surface, polymer remains attached to the nanoparticle surface, which cannot move freely. It is expected that the top layer close to the continuous oil phase is first polymerized to become a skin layer, which is harder than the inner layer close to the nanoparticle due to the gradient in cross-linking reaction. Additionally, NIPAM polymer is known to be highly sensitive to temperature changes.56 In our UV curing system, the chamber temperature was approximately 65 °C during the exposure. Such temperature is higher than its low critical solution temperature (LCST of NIPAM polymer) of 32 °C, causing the NIPAM polymer to become hydrophobic, expelling water.57 This leads to the drastic shrinkage of the formed skin layer. When the polymer is strongly attached to the nanoparticle surface, no detachment takes place, and the shrinkage of NIPAM polymer produces an in-plane tensile stress in the skin layer. The elastic modulus of the NIPAM polymer layer as a skin layer increases by an order of magnitude compared to that of the inner soft layer.57 This difference in the elastic modulus between the two layers essentially results in the formation of nanowrinkles. On the other hand, due to the gradient in the chemical potential during the cross-linking reaction, the monomers/oligomers tend to diffuse toward the hard skin layer, thus resulting in a swelling tendency. Satisfaction of further cross-linking promotes monomers/oligomers to diffuse deeper into the skin, which as a result slowly changes the state of stress of the skin from tensile to compressive. Therefore, a competition between the tensile stress (caused by the shrinkage of the skin layer from chemical reaction) and the compressive stress (resulted from the swelling) is built (Figure 4c). In order to release the stresses, an out-of-plane deformation accompanied by nanowrinkles occurs (Figure 4d).

On a flat surface, the wavelength of nanowrinkles can be characterized by the equation of \( \lambda \approx 2\pi h (E_s/E_B)^{1/3} \), where \( h \) is the thickness of the skin layer, \( E_s \) and \( E_B \) are the Young’s modulus of the skin hard and inner soft layers, respectively.58,59 This indicates that the characteristics of the nanowrinkles can be
Figure 6. HR-SEM images of films prepared by assembly of (a) PS-co-PAA nanoparticles (scale bar: 200 nm), (b) PS-co-PAA@MS(T) (scale bar: 1 μm), and (c) f-PS@MS (scale bar: 1 μm). Inset HR-SEM images in (b) and (c) show the amplified nanopattern surfaces (scale bar: 200 nm). Inset optical images in (a–c) show the CA measurements of 2 μL water drops sitting on the corresponding films (scale bars: 200 μm). The schematic diagrams on the bottom rows show corresponding geometries and parameters of the three films. (d) A water drop standing on a film covered by the three tier f-PS@MSs, tilted at different angles (scale bars: 1 mm).

We conclude from the above investigation that the surface topology of hierarchical microspheres can be precisely controlled by changing the fraction of carboxylic groups on the PS nanoparticle surface, as shown in Figure 5. With a high density of carboxylic groups on the PS nanoparticle surface, the produced wrinkles show high spatial frequency and roughness. Bioinspired “Gecko-like” and “Rose Petal-like” Surfaces from Self-Assembly of Two- and Three-Tier Microspheres. Surfaces making use of versatile topological structures have attracted a lot of attention due to their wide applicability in waterproofing, self-cleaning, antifouling, and anti-icing. Surface wettability is strongly dependent on both the tailored structure and chemical modification, thereby tuning the adhesion and friction forces between fluids and solid surfaces. We investigated the effect of the surface topography on the surface wettability of different films composed of closely packed PS nanoparticles (PS-co-PAA), two-tier hierarchical microspheres constructed by PS nanoparticles (PS-co-PAA@MS(T)), or three-tier hierarchical microspheres constructed by PS nanoparticles covered with nanowrinkles (f-PS@MS). In Figure 6a, a film of closely packed PS nanoparticles was assembled by using PS-co-PAA nanoparticles (250 nm in diameter). In Figure 6b (see also AFM images in Figure S5), the two-tier hierarchical microspheres (PS-co-PAA@MS(T)) were generated from the same PS-co-PAA nanoparticles using the droplet-based microfluidics approach with evaporation-induced assembly and solidification. In Figure 6c (see also AFM images in Figure 2), the three-tier hierarchical microspheres (f-PS@MS) were produced from f-PS nanoparticles of 250 nm in diameter using the same method with photopolymerization-induced solidification. The sub-30 nm nanowrinkles on f-PS@MS led to significant enhancement of surface hydrophobicity, suggested by the increase in water contact angle (CA) measurements.

Physical equations have been used to express the relationship between the surface roughness and wettability, typically the Wenzel and Cassie-Baxter equations. In the Wenzel state, the water drop totally wets the substrate, forming a continuous three-phase contact line (TCL), resulting in a high adhesive force (high CA hysteresis). In contrast, in the Cassie-Baxter state, air is trapped at the interface between the water drop and the substrate, forming a nonwetting contact mode and a discontinuous and unstable TCL, resulting in low adhesion (low CA hysteresis). The Wenzel and Cassie-Baxter states are considered as ideal models for a drop sitting on a superhydrophobic surface. However, practically, there are several intermediate states such as the “gecko” and the “rose petal” states, existing on both natural and artificial hierarchical structures. In the “gecko” state, a water drop is sucked into the top part of nanoscale capillaries, resulting in sealed air pockets trapped in the bottom of the nanoscale spaces. This leads to the formation of a negative pressure and thus produces a high adhesive force. In the “rose petal” state, a water drop only partially wets the textured structure of a superhydrophobic surface. When a water drop sits on a “rose petal” surface, the water impregnates only into the large-scale micropapillae of the petal without filling the nanofolds on top of the micropapillae.
thus leaving air trapped at the nanofolds. Such a wetting state enables the water drop to be tightly adhered to the rose petal surface, even when it is turned upside-down.66

The films prepared from PS-co-PAA nanoparticles, PS-co-PAA@MS(T), and f-PS@MS showed CAs of 25°, 130°, 145° in Figure 6a–c, respectively. The surface of assembled PS-co-PAA nanoparticles showed a Wenzel state with the water drop wetting the surface and forming a continuous TCL, as shown in Figure 6a. This 2D film was featured with high-density nanogaps with 30 ± 10 nm distance among neighboring PS-co-PAA nanoparticles. Such a structure leads to much more hydrophobic comparison in a flat polystyrene film (CA of ~90°).66

Figure 6b shows a surface composed of the well-patterned two-tier microspheres of PS-co-PAA@MS(T), including the micropatterns from assembled microspheres as the frames (analogy to the setae of gecko’s foot) and the nanopatterns of closely packed nanoparticles (analogy to the spatulae on setae), which shows a “gecko” state.68 From the AFM image and the corresponding dimensions of the two-tier microspheres (PS-co-PAA@MS(T)) in Figure S5, the height of the nanogaps among nanoparticles was approximately 100 nm and the minimum gap distance between two nanoparticles was about 15 ± 10 nm.

The surface of closely packed three-tier hierarchical microspheres of f-PS@MS exhibited a “rose petal” state, as presented in Figures 6c, with higher CA and high adhesion to water. Hierarchical structures featured with micropapillae and nanofolds are known to exist on the petal surfaces of red roses.6 The produced film fabricated by three-tier microspheres was similar to the “rose petal” surface, where the three-tier microsphere units serve as the micropapillae, and nanowrinkles on the microsphere are like nanofolds on every micropapillae. Compared to the surface of assembled two-tier microspheres in Figure 6b, the three-tier microsphere assembled surface showed higher water CA of 145° as well as high adhesion to water. This is mainly attributed to the enhanced roughness and spatial frequency from the multiple-tier of nanostructures and microstructures. Figure 6d shows that a water drop stably remained on the surface of assembled three-tier f-PS@MS microspheres (Figure 6c), even when it is turned 180° upside-down (Video S1).

Overall, these three states of surface wettability are mainly determined by the physical micro- and nanoengineered structures. In this way, by using the microfluidic platform, we could create the microunits with controllable nanostructures for constructing surfaces with variable wettability. With the combination of nanoparticles and polymeric materials, the multilayer nano- and microstructures could be manipulated. Further modification of the film surfaces with tunable chemical compositions is expected to be able to better regulate the surface wettability in a better controllable manner.

CONCLUSION

In summary, we report a robust and facile method for the rapid fabrication of three-tier hierarchical microspheres featuring nanostructures using a droplet-based microfluidic platform, which has the advantages of precise control over particle size, flexible tunability of materials, and easy integration with other techniques. These hierarchical microspheres with multiple structures could be applied as constructable units to build up bioinspired surfaces by self-assembly on various substrates. Both the microsphere size and generation frequency can be controlled by the microfluidic device design and flow dynamics of oil and water phases. The nanometer-scale patterns on the microsphere stem from the self-assembly of colloidal nanoparticles. The nanowrinkles with sizes in the range of tens of nanometers are generated from the gradient polymerization-induced stress mismatch between the soft NIPAM monomers/oligomers that are strongly bound to PS surface via hydrogen bonds and the PNIPAM (polymerized NIPAM) hard skin layer. The roughness and spatial frequency of the nanowrinkles could be tailored by the density of carboxylic groups at the PS nanoparticle surfaces. Due to the morphology in three spatial levels at the nano- and microscale, these microspheres show high roughness and spatial frequency. Surfaces made from self-assembly of the two- and three-tier microsphere exhibit “gecko” and “rose petal” wetting states, showing hydrophobicity with CAs of 130 and 145°, respectively, but high water adhesion to water by holding a drop without falling off when the film is tilted. With implementation of chemical control over the microspheres-nanoparticles-nanowrinkles, more applications could be expected for this reported method for creating 3D three-tier hierarchical structures as catalytic support, gas sensing units, and sample preconcentration materials.

METHODS

Materials. N-isopropylacrylamide (NIPAM, purity ≥99%), N,N-methylene bis(acrylamide) (MBA, purity ≥99%), n-hexadecane (purity 99%), sorbitane monoooleate sorbitan oleate (Span 80), and 2,2-dioctoxyacetophenone (DEAP, purity ≥98%) were purchased from Sigma-Aldrich (The Netherlands). Both bare polystyrene nanoparticles (bare-PS nanoparticles, d = 250 ± 10 nm) and copolymerized polystyrene nanoparticles with two different acrylic acid (AA) molar fractions (10% PS-co-PAA, d = 250 ± 10 nm nanoparticles, 18.5% PS-co-PAA, d = 250 ± 10 nm) were provided by Nanjing Nanorainbow Biotechnology Co., Ltd. company (Nanjing, China). The PS-co-PA nanoparticles were synthesized from random polymerization by the monomer of styrene and acrylic acid with different contents via an soap-free emulsion polymerization method. The chemical structure of these different polystyrene nanoparticles used in this work is shown in Figure S3a. The fluorescent polystyrene nanoparticles with carboxylic functionalized surface (defined as f-PS nanoparticles, d = 250 ± 50 nm) were purchased from Wuhan Jiayuan Quantumdots Co. Ltd. company (Wuhan, China). The synthetic route and chemical structure of f-PS nanoparticles are shown in Figure S3b. Hydrazine-modified polystyrene nanoparticles were treated further with the succinic anhydride, yielding the carboxylic-functionalized f-PS nanoparticles.67 The HR-SEM images of the morphological surface of bare-PS, PS-co-PAA, and f-PS nanoparticles are shown in Figures S3c–f. Polystyrene nanoparticles were all dispersed in deionized water without any additions before using to generate emulsion droplets.

Fabrication of the Microfluidic Device. The polydimethylsiloxane (PDMS) microfluidic device with flow-focusing geometry was fabricated using the standard soft lithography technique.6 The detailed fabrication process of this microfluidic device was presented in our previous work.55

Construction of Hierarchical Microspheres. Two immiscible phases were used to produce the highly uniform emulsion droplets through the flow-focusing microfluidic device. The water phase (dispersed phase) was a suspension of different polystyrene nanoparticles with a certain amount of monomer (NIPAM) and cross-linker (MBA). The oil phase (continuous phase) was n-hexadecane contained Span 80 (as the surfactant to stabilize the generated emulsion droplets) and DEAP (as the photoinitiator). The water phase was sheared and broken into monodispersed emulsion droplets by the oil phase in the flow focusing area of the microfluidic device. Produced monodispersed droplets containing numbers of nanoparticles served as a template to induce the self-assembly of nanoparticles on the droplet surface driven by surface energy minimization and stabilized by the electrostatic repulsion. As the time evolved, the droplets were featured with close-
packed nanoparticles, which were then solidified upon UV irradiation (IntelliRay-600, 365 nm, 600 W, 120 mW cm\(^{-2}\), Uvitrон International, Inc., Massachusetts) for 300 s through photopolymerization, yielding solid hierarchical microspheres (denoted as MS). In addition, the thermal evaporation method was also used to yield solid microspheres (denoted as MS(\(T\)) as a control approach. Herein, the composition of hierarchical microcapsules can be precisely and flexibly tuned, which are attributed to the advantages of microfluidic techniques.

**Characterization. Optical Microscope.** The generation process of emulsion droplets was monitored and recorded by using an inverted optical microscope (Olympus IX2, Tokyo, Japan) equipped with a high-speed camera (Phantom, MIRO MIIO, Vision Research Inc., Wayne County, NC, USA) in bright-field mode. The fluorescent images were taken by an inverted optical microscope (Olympus IX73, Tokyo, Japan) equipped with a digital camera (Olympus DP73, Tokyo, Japan) using the 365 nm excitation mode.

**Scanning Electron Microscope.** Fabricated hierarchical microsphere suspension in ethanol was dripped onto a clean piece of silicon substrate and dried naturally at room temperature. A thin platinum layer was coated on the exposed surface using a coating machine (Quorus, Q50T ES, Quorum Technologies Ltd., Ashford, England) prior to the HR-SEM examination. The morphology of these microspheres was characterized by FE-SEM (ZEISS ultra 55, Carl Zeiss, Germany) using either the secondary electron signal or in-lens signal was coated on the exposed surface using a coating machine using the 365 nm excitation mode.

**Atomic Force Microscope.** Dimensions of nanoparticles on the surface of two-tier/three-tier hierarchical microspheres were measured by using an atomic force microscope (AFM) (Bruker Fast Scan, Bruker Corporation, Billerica, United States) equipped with a Fast Scan scanner for high-resolution images. The contact-mode AFM with a tip radius of 2 nm and scanning frequency in the range of 0.99–1.99 Hz was used to collect the topological surface.

**Fourier-Transform Infrared Spectroscopy.** Poly(styrene) nanoparticles (PS-co-PAA and f-PS nanoparticles) were analyzed by using a Fourier-transform infrared spectroscopy (FT-IR) under the transmittance mode (PerkinElmer Spectrum 2000, USA) in the region of 400–4000 cm\(^{-1}\). Before each measurement, the sample was vacuumed and dried in an oven overnight. The dried samples were mixed with KBr by grinding and compressed into a round transparent sheet for FT-IR analysis. The adsorption spectra of PS-co-PAA nanoparticles, f-PS nanoparticles, and hierarchical microspheres (f-PS@MS) made from f-PS nanoparticles analyzed by FT-IR are shown in Figure S6a1–c1, respectively. Figure S6a2–c2 shows the locally amplified transmission spectra in the wavenumber of 1500–1800 cm\(^{-1}\) corresponding to Figure S6a1–c1.

The transmittance peaks at 3068, 3026, 1596, 1492, 1448, 756, and 696 cm\(^{-1}\) correspond to the phenyl (Ph) group; the peaks at 2920 and 2846 cm\(^{-1}\) correspond to the methylene and methenyl groups, respectively; and the peaks at 1072 and 1024 cm\(^{-1}\) correspond to the C–C and C–Ph bonds, respectively (see spectra in Figure S6a1–c1). This indicated the presence of the bone polymer (polystyrene). Moreover, each spectrum showed its own characteristic peaks, demonstrating different functional groups produced on the corresponding particles. As shown in Figure S6a1–a2, the peaks located at 3448 and 1701 cm\(^{-1}\) were resulted from stretching vibrations of hydroxyl group and carbonyl group C=O, respectively. 180 indicated the existence of carboxylic group on PS-co-PAA nanoparticles. Figure S6b1 shows the presence of amide group and carboxylic group on the specifically carboxylic functionalized nanoparticles (f-PS), resulting from the N–H stretch at 3431 cm\(^{-1}\), the C=O stretch from amide group at 1641 cm\(^{-1}\), and the weak peak of O–H stretch and C=O stretch from carboxylic group located at 3730 and 1745 cm\(^{-1}\), respectively. The produced three-tier hierarchical microspheres (f-PS@MS) with nanowrinkles were also analyzed by using FT-IR (see Figure S6c1). The broad band at 3464 cm\(^{-1}\) is attributed to the N–H stretching vibration, and the strong peak at 1647 cm\(^{-1}\) is due to C=O stretching, demonstrating all typical absorption peaks of NIPAM polymer. 19 The peak located at 1101 cm\(^{-1}\) indicated the stretch of the C–N group.

**X-ray Photoelectron Spectroscopy.** X-ray photoelectron spectroscopy (XPS) was utilized for surface analysis, by using an Omicron Nanotechnology GmbH (Oxford Instruments) with a photon energy of 1486.7 eV (Al Kα X-ray source) at a scanning step size of 0.1 eV. Before the measurement, the samples were dried and transferred onto a clean piece of carbon film. XPS spectra were obtained with a spot size of 200 μm. High-resolution scans of individual elements and the survey spectra were measured at a pass energy of 20 and 200 eV, respectively. The scans were averaged over 10 sweeps.

The XPS spectra of the PS-co-PAA and f-PS nanoparticles are shown in Figure S7a,d, respectively. Obviously, the PS-co-PAA nanoparticles were composed of C and O elements without N element. The spectrum of f-PS nanoparticles showed the presence of N element, which is consistent with the chemical structure and component of the as-prepared f-PS nanoparticles, as shown in Figure S3. The high-resolution spectrum of C 1s from the PS-co-PAA nanoparticles (Figure S7b) showed three peaks at 285.2, 286.7, and 289.2 eV for C–C, C–O, and C(=O)O, respectively. The high-resolution spectrum of O 1s (Figure S7c) revealed the presence of C–O bonds (532.5 eV) and C≡O bonds (533.7 eV). Moreover, the high-resolution spectrum of C 1s from f-PS nanoparticles (Figure S7e) showed three bands located at C–C bonds (284.6 eV), C–N bonds (285.7 eV), and C≡O (287.9 eV), respectively. The peaks at C–O (531.1 eV) and C≡O (532.7 eV) were also presented in the high-resolution spectrum of O 1s from f-PS nanoparticles (Figure S7f).

**Preparation of Bioinspired Surfaces.** The hierarchical microsphere suspension (dispersed in deionized water) was pipetted onto a piece of silicon wafer cleaned by using oxygen plasma. Due to hydrophobicity, the microspheres floated on the water surface and started to assemble at room temperature naturally as water evaporated. The obtained films with multilayer patterned microspheres were then used to study the wetting performance (Figure S8).

**Contact Angle Measurement.** One water drop with a volume of 2 μL was dripped gently on the as-prepared substrate. The substrate was covered with a layer of produced hierarchical microspheres. An international tensil meter OCA 15 Pro (Dataphysics, Germany) was used to measure the contact angle at ambient temperature. The Laplace–Young fitting method was used to measure the value of the contact angle. Every substrate was measured for five times at different positions to obtain the average contact angle.

**ASSOCIATED CONTENT**

† Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsnano.9b00245.

Estimation of the volume shrinkage fraction of droplets, the surface-to-volume ratio of a f-PS@MS, and the density of nanogaps, the schematic of different PS nanoparticles and the corresponding HR-SEM images, HR-SEM images of hierarchical microspheres made from PS-co-PAA nanoparticles, and PS-co-PAA nanoparticles, and XPS images of PS-co-PAA@MS(T), and FT-IR spectra, XPS spectra (PDF)

Video S1: Demonstration of one 2 μL water drop sitting on the surface made from assembled three-tier hierarchical microspheres (MPG)

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Notes
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