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Anchoring of histidine-tagged proteins to molecular printboards: self-assembly, thermodynamic modeling and patterning

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Dedicated to Professor David N. Reinhoudt on the occasion of his 65th birthday

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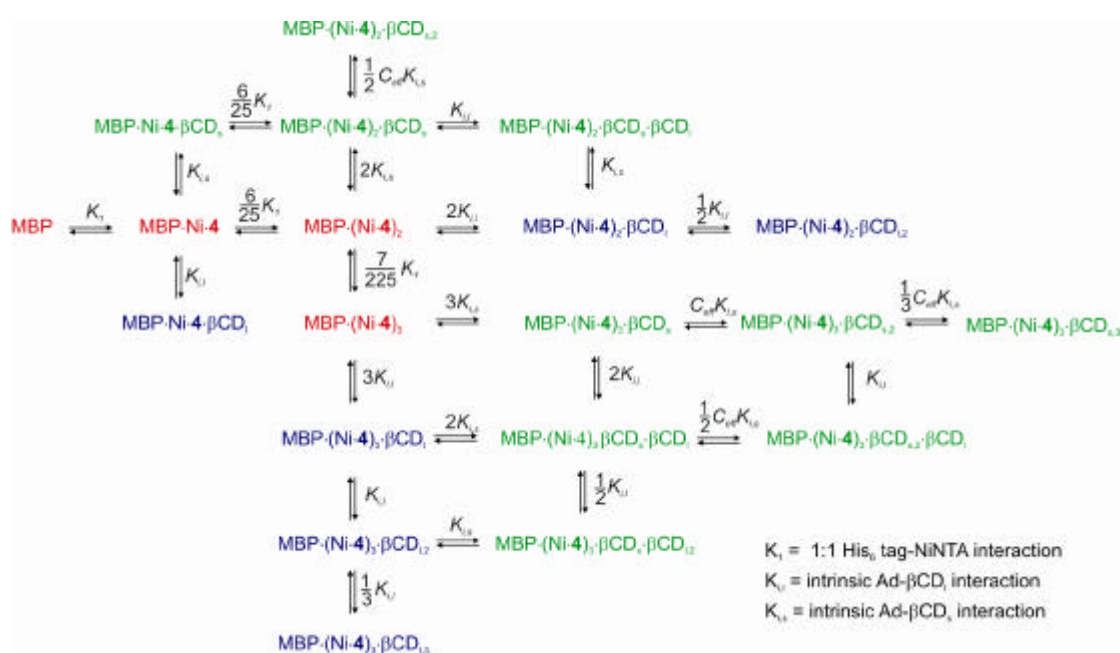
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Modeling of the binding of His₆-MBP to the molecular printboard

The binding of His₆-MBP to **bCD** SAMs via Ni•4 can be monovalent, divalent, or trivalent. In Scheme 2 all possible equilibria are presented, assuming the Ni•4 complex forms completely, as discussed in the main text. At **bCD** SAMs, all His₆-MBP units complexed to one Ni•4 will behave as monovalent guests, binding to surface-confined **bCD** (**bCD**_s) in a similar fashion as to **bCD** in solution (**bCD**_l). For His₆-MBP units that are bound via two or three Ni•4 complexes, the binding to **bCD** SAMs is governed by an effective concentration term (C_{eff}), which is the driving force for the formation of multivalent complexes at **bCD** SAMs. It has to be noted, that the effective concentration represents the probability that an unused guest site finds a complementary host site and thus incorporates all entropic multivalency factors, including e.g. entropy changes due to conformational losses between the guest sites. For the system at hand, the effective concentration concept provides a rigorous quantitative description of the equilibria involved.^[1,2]

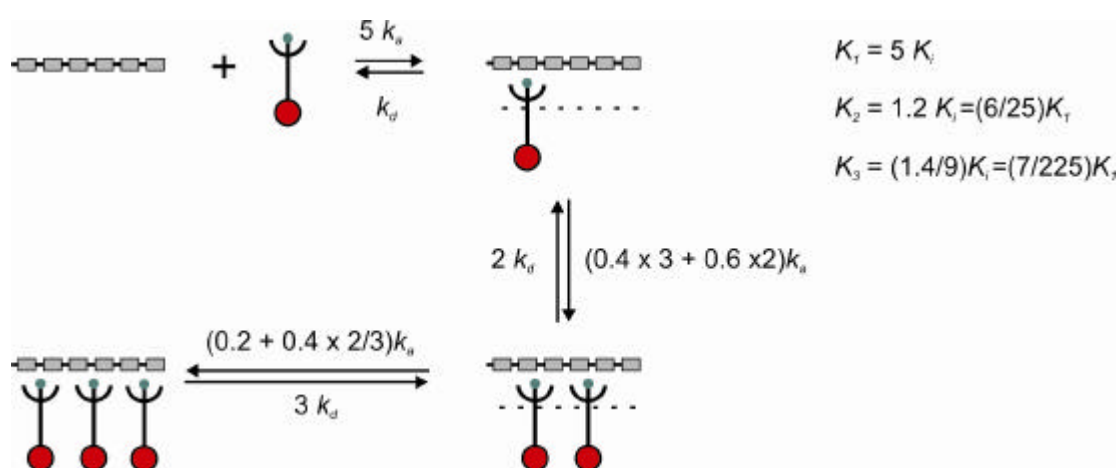


Scheme 2. Equilibria for all species (solution and surface) for the attachment of His₆-MBP at the molecular printboard (charges are omitted for clarity). Subsequent complexation steps of Ni•4 to MBP are shown in red, and all surface species are given in green.

A general description is given for the multivalent binding of His₆-MBP•(Ni•4)_x (x=1-3) to the molecular printboard. The stepwise adsorption of e.g. His₆-MBP•(Ni•4)₃ to the surface involves an intermolecular adsorption step and two intramolecular binding steps, the latter of which are both governed by C_{eff} . All solution and surface species of MBP are shown in Scheme 1. All intrinsic stability constants for bCD_l and bCD_s are assumed equal for all steps given in Scheme 2.^[1]

Similar to the binding studies described before,^[1,2] SPR titrations performed for the binding of His₆-MBP to bCD SAMs in the presence of Ni•4 are fitted here, yielding K_1 values for the His₆ tag-Ni•4 interaction, while the intrinsic binding constant of an adamantyl guest to bCD in solution and the intrinsic binding constant of an adamantyl guest to a surface-confined bCD cavity, $K_{i,l}$ and $K_{i,s}$, respectively, are fixed to the values determined by ITC and SPR.

The statistical factors relating K_2 and K_3 for additional Ni•4 were determined by noting that: (i) Ni•4 binds to two neighboring histidines, (ii) binding Ni•4 to His₆-MBP is 5 times as likely as binding to a His₂ unit (which is the intrinsic interaction motif in this case), (iii) there are 2 or 3 His₂ sites free for interaction of a second Ni•4 to MBP•Ni•4 in 60% and 40% of the MBP•Ni•4 complexes, respectively, and (iv) only for 46.7% of the MBP•(Ni•4)₂ complexes there is an additional free His₂ site available for a third Ni•4 unit (See Scheme 3). This leads to the prefactors of $K_2 = \frac{6}{25}K_1$ and $K_3 = \frac{7}{225}K_1$ as given in Scheme 2.



Scheme 3. The statistical factors relating K_1 , K_2 and K_3 to K_i .

Since all measurements have been performed at pH = 7.5, Ni•4 is always formed completely (see main text). The mass balances that can be constructed based on Scheme 2 are the following (charges are omitted for clarity):

$$\begin{aligned}
[\text{MBP}]_{\text{tot}} = & [\text{MBP}]_{\text{free}} + [\text{MBP}\cdot\text{Ni}\cdot\mathbf{4}] + [\text{MBP}\cdot\text{Ni}\cdot\mathbf{4}\cdot\beta\text{CD}_1] + [\text{MBP}\cdot\text{Ni}\cdot\mathbf{4}\cdot\mathbf{bCD}_s] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_2] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_2\cdot\mathbf{bCD}_s] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_2\cdot(\mathbf{bCD}_s)_2] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_2\cdot\mathbf{bCD}_1] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_2\cdot(\mathbf{bCD}_1)_2] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_2\cdot\mathbf{bCD}_s\beta\text{CD}_1] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot(\mathbf{bCD}_s)_2] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot\mathbf{bCD}_1] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot(\mathbf{bCD}_1)_2] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot(\mathbf{bCD}_1)_3] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot\mathbf{bCD}_s] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot\mathbf{bCD}_s\beta\text{CD}_1] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot\mathbf{bCD}_s(\mathbf{bCD}_1)_2] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot(\mathbf{bCD}_s)_2\mathbf{bCD}_1] + [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot(\mathbf{bCD}_s)_2] + \\
& [\text{MBP}\cdot(\text{Ni}\cdot\mathbf{4})_3\cdot(\mathbf{bCD}_s)_3]
\end{aligned} \tag{1}$$

$$\begin{aligned}
[4]_{\text{tot}} = & [Ni\bullet4]_{\text{free}} + [Ni\bullet4\bullet bCD_1] + [Ni\bullet4\bullet bCD_s] + [MBP\bullet Ni\bullet4] + \\
& [MBP\bullet Ni\bullet4\bullet bCD_s] + [MBP\bullet Ni\bullet4\bullet bCD_1] + 2([MBP\bullet(Ni\bullet4)_2] + \\
& [MBP\bullet(Ni\bullet4)_2\bullet bCD_s] + [MBP\bullet(Ni\bullet4)_2\bullet(bCD_s)_2] + [MBP\bullet(Ni\bullet4)_2\bullet bCD_1] + \\
& [MBP\bullet(Ni\bullet4)_2\bullet(bCD_1)_2] + [MBP\bullet(Ni\bullet4)_2\bullet bCD_1\bullet bCD_s]) + \\
& 3([MBP\bullet(Ni\bullet4)_3] + [MBP\bullet(Ni\bullet4)_3\bullet bCD_s] + [MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_2] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_3] + [MBP\bullet(Ni\bullet4)_3\bullet bCD_1] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet(bCD_1)_2] + [MBP\bullet(Ni\bullet4)_3\bullet(bCD_1)_3] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet bCD_s\bullet bCD_1] + [MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_2\bullet bCD_1] + \\
& [MBP(Ni\bullet4)_3\bullet bCD_s\bullet(bCD_1)_2]) \tag{2}
\end{aligned}$$

$$[3]_{\text{tot}} = [3]_{\text{free}} + [3\bullet bCD_1] + [3\bullet bCD_s] \tag{3}$$

$$\begin{aligned}
[bCD_s]_{\text{tot}} = & [bCD_s]_{\text{free}} + [MBP\bullet Ni\bullet4\bullet bCD_s] + [MBP\bullet(Ni\bullet4)_2\bullet bCD_s] + \\
& [MBP\bullet(Ni\bullet4)_2\bullet bCD_s\bullet bCD_1] + [MBP\bullet(Ni\bullet4)_3\bullet bCD_s] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet bCD_s\bullet bCD_1] + [MBP\bullet(Ni\bullet4)_3\bullet bCD_s\bullet(bCD_1)_2] + \\
& 2([MBP\bullet(Ni\bullet4)_2\bullet(bCD_s)_2] + [MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_2] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_2\bullet bCD_1]) + 3[MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_3] + [3\bullet bCD_s] + \\
& [Ni\bullet4\bullet bCD_s] \tag{4}
\end{aligned}$$

$$\begin{aligned}
[bCD_1]_{\text{tot}} = & [bCD_1]_{\text{free}} + [MBP\bullet(Ni\bullet4)_2\bullet bCD_s\bullet bCD_1] + [MBP\bullet(Ni\bullet4)_2\bullet bCD_1] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet bCD_1] + [MBP\bullet(Ni\bullet4)_3\bullet bCD_s\bullet bCD_1] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet(bCD_s)_2\bullet bCD_1] + 2([MBP\bullet(Ni\bullet4)_2\bullet(bCD_1)_2] + \\
& [MBP\bullet(Ni\bullet4)_3\bullet(bCD_1)_2] + [MBP\bullet(Ni\bullet4)_3\bullet bCD_s\bullet(bCD_1)_2]) + \\
& 3[MBP\bullet(Ni\bullet4)_3\bullet(bCD_1)_3] + [3\bullet bCD_1] + [Ni\bullet4\bullet bCD_1] + \\
& [MBP\bullet Ni\bullet4\bullet bCD_1] \tag{5}
\end{aligned}$$

Species involving bCD_s are expressed in volume concentrations.^[1] The binding of the divalent $MBP\bullet(Ni\bullet4)_2$ and trivalent $MBP\bullet(Ni\bullet4)_3$ to bCD_1 involves statistical factors (Scheme 2) arising from the probabilities for binding relative to the monovalent species, in this case according to a normal 1:3 complexation sequence.

The binding constants for first intermolecular binding events of the divalent and trivalent species at the surface are:

$$K = \frac{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_2 \cdot \mathbf{bCD}_s]}{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_2][\mathbf{bCD}_s]} = 2K_{i,s} \quad (6)$$

$$K = \frac{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_3 \cdot \mathbf{bCD}_s]}{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_3][\mathbf{bCD}_s]} = 3K_{i,s} \quad (7)$$

The second, intramolecular, binding event for the di- and trivalent species, and third, for the trivalent species (equations 9, 10, and 11) are governed by an effective concentration term, which is defined as given in equation 8.^[1,2] The effective concentration is given by multiplying the maximum effective concentration, $C_{eff,max}$, which is the number of accessible host sites in the probing volume, with the fraction of free host sites at the surface.

$$C_{eff} = C_{eff,max} \frac{[\mathbf{bCD}_s]}{[\mathbf{bCD}_s]_{tot}} \quad (8)$$

$$K = \frac{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_2 \cdot (\mathbf{bCD}_s)_2]}{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_2 \cdot \mathbf{bCD}_s][\mathbf{bCD}_s]} = \frac{1}{2} C_{eff} K_{i,s} \quad (9)$$

$$K = \frac{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_3 \cdot (\mathbf{bCD}_s)_2]}{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_3 \cdot \mathbf{bCD}_s][\mathbf{bCD}_s]} = C_{eff} K_{i,s} \quad (10)$$

$$K = \frac{[\text{MPB} \cdot (\text{Ni} \cdot \mathbf{4})_3 \cdot (\mathbf{bCD}_s)_3]}{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})_3 \cdot (\mathbf{bCD}_s)_2][\mathbf{bCD}_s]} = \frac{1}{3} C_{eff} K_{i,s} \quad (11)$$

Since the SPR experiments were performed in a flow system, all solutions species concentrations can be calculated from simplified forms of equations 1-3 and 5. After numerical optimization of these equations, the values obtained for the solution species concentrations were used in the full equations 1-5 for calculations of the surface species.

Substitution of the equilibrium constant definitions into the mass balances for $[\text{MBP}]_{\text{tot}}$, $[\text{bcd}_s]_{\text{tot}}$, $[\text{bcd}_l]_{\text{tot}}$, $[\mathbf{3}]_{\text{tot}}$, and $[\mathbf{4}]_{\text{tot}}$ (equations 1-5) provides a set of numerically solvable equations with $[\text{MBP}]$, $[\text{bcd}_s]$, $[\text{bcd}_l]$, $[\mathbf{3}]$, and $[\mathbf{4}]$ as the variables.

Starting from an initial estimate for K_I (defined as: $K_I = \frac{[\text{MBP} \cdot (\text{Ni} \cdot \mathbf{4})]}{[\text{MBP}][\text{Ni} \cdot \mathbf{4}]}$) using fixed

values for $C_{\text{eff,max}}$ (0.1 M) and the other stability constants, this set of equations is solved numerically using a Simplex algorithm in a spreadsheet approach.^[3] When fitting SPR data, K_I is optimized in a least-squares optimization routine, assuming that the SPR response (intensity) is linearly dependent on the coverages of MBP, $\mathbf{3}$, and $\mathbf{4}$ adsorbed to the **bcd** SAM, regardless of the type of species. The maximum intensity (I_{max} of MBP) is then optimized as an independent fitting parameter as well while those of $\mathbf{3}$ and $\mathbf{4}$ were determined by independent SPR measurements.

Based on Scheme 1, the overall stability constant for $\text{His}_6\text{-MBP} \cdot (\text{Ni} \cdot \mathbf{4})_3 \cdot (\text{bcd}_s)_3$ can be given by equation 12:

$$K = (K_{i,s})^3 K_I K_2 K_3 C_{\text{eff}}^2 \cdot [\text{Ni} \cdot \mathbf{4}]^3 \quad (12)$$

Assuming $K_{i,s} = 1.2 \times 10^4 \text{ M}^{-1}$, $K_I = 7.8 \times 10^3 \text{ M}^{-1}$ ($K_2 = \frac{6}{25} K_I$ and $K_3 = \frac{7}{225} K_I$),

$C_{\text{eff}} = C_{\text{eff,max}} = 0.1 \text{ M}$ (at relatively low coverages), and $[\text{Ni} \cdot \mathbf{4}] = 1 \text{ } \mu\text{M}$, an apparent conditional binding constant of $\sim 10^5 \text{ M}^{-1}$ can be estimated.

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