# <sup>13</sup>C N.m.r. Investigation on the First and Second Nitrogen Protonation in the Diazanaphthalenes

Peter van de Weijer,\* Dirk M. W. van den Ham and Douwe van der Meer

Chemical Physics Laboratory, Twente University of Technology, Enschede, the Netherlands

The  $^{13}$ C chemical shifts of the diazanaphthalenes have been recorded as a function of the pH value, providing classical titration curves. From these curves the p $K_1$  and p $K_2$  values have been determined taking into account the activity coefficients. The changes in  $^{13}$ C chemical shift under the influence of nitrogen protonation ( $\Delta\delta$ ) can be described by two protonation parameter sets.

## INTRODUCTION

Recently we have reported a <sup>13</sup>C n.m.r. investigation on the site of protonation in the diazanaphthalenes. <sup>1</sup> This group of N-heterocycles consists of ten compounds: cinnoline (1,2-diazanaphthalene, 1), quinazoline (1,3-diazanaphthalene, 2), quinoxaline (1,4-diazanaphthalene, 3), 1,5-naphthyridine (1,5-diazanaphthalene, 4), 1,6-naphthyridine (1,6-diazanaphthalene, 5), 1,7-naphthyridine (1,7-diazanaphthalene, 6), 1,8-naphthyridine (1,8-diazanaphthalene, 7), phthalazine (2,3-diazanaphthalene, 8), 2,6-naphthyridine (2,6-diazanaphthalene, 9) and 2,7-naphthyridine (2,7-diazanaphthalene, 10).

In order to establish the site of protonation we recorded the pH dependence of the chemical shift of each separate carbon atom in all isomers.

From the titration curves thus obtained we simultaneously derived the pK values of the first nitrogen protonation  $(pK_1)$  by application of the Henderson-Hasselbach equation.<sup>2</sup> These  $pK_1$  values correlated very well with those values from the literature that were derived by conventional titration methods. However, the  $pK_1$  values determined with the n.m.r. technique proved to be systematically smaller than the literature values. The same has been noticed for other aza-aromatics by Breitmaier and Spohn.<sup>3</sup>

The most obvious explanation of this systematic deviation is that in the  $^{13}$ C n.m.r. experiment the concentration of the titrant is much higher than in a normal titration procedure. As a consequence, the activity coefficients of the participating species will deviate seriously from unity. In  $^{13}$ C n.m.r. determinations of the second nitrogen protonation constants  $(pK_2)$  this factor gains even more importance because of the still higher ionic strength of the solutions involved.

In this contribution we apply a procedure that takes into account these activity coefficients. Since this method proved to be successful for the  $pK_1$  values a similar approach was used for the determination of the  $pK_2$  values.

While determining the pK values the changes in the  $^{13}$ C chemical shifts could be followed. These changes upon first and second nitrogen protonation ( $\Delta\delta_1$  and  $\Delta\delta_2$ , respectively) can be described by two protonation parameter sets. <sup>1</sup>

### **pK DETERMINATION**

### Method

In a previous study<sup>1</sup> we made the assumption that the ratio of activity coefficients  $f_{\rm RH}^+/f_{\rm R}$  is equal to unity. This holds for dilute solutions. However, for the <sup>13</sup>C n.m.r. experiment relatively high concentrations are desirable, causing  $f_{\rm RH}^+/f_{\rm R}$  to deviate from unity. By taking into account this deviation we intend to establish better agreement between our p $K_1$  values and the ones described in the literature.

The  $pK_1$  value of the equilibrium

$$R + H^+ \leftrightharpoons RH^+$$

may be described by the Henderson-Hasselbach equation:<sup>2</sup>

$$pK_1 = pH + log[(f_{RH^+} \cdot c_{RH^+})/(f_R \cdot c_R)]$$

where c is the concentration.

The ratio  $I = c_{RH^+}/c_R$  has been determined by the  $^{13}$ C n.m.r. technique as  $(\delta_1 - \delta)/(\delta - \delta_2)$  where  $\delta_1$  is the chemical shift of R,  $\delta_2$  is the chemical shift of RH<sup>+</sup> and  $\delta$  is the chemical shift of a mixture of R and RH<sup>+</sup> at a certain pH value. Although the  $\delta_1$  and  $\delta_2$  values are slightly solvent and counter ion dependent we assumed them to be constant. It has been demonstrated that this is a reasonable assumption since the correlation between log I and pH is excellent (correlation coefficient > 0.999).

Hammett has demonstrated<sup>4</sup> that for structurally similar compounds the ratio  $f_{\rm RH^+}/f_{\rm R}$  is a constant at a certain fixed pH value. The diazanaphthalenes seem to satisfy this condition very well. Consequently the difference between the p $K_1$  values of two structurally similar compounds at the same pH value may be

<sup>\*</sup> Author to whom correspondence should be addressed.

<sup>©</sup> Heyden & Son Ltd, 1977

determined by

$$\Delta p K_1 = \Delta \log (c_{RH^+}/c_R) = \Delta \log I$$

This equation provided us with the relative  $pK_1$  values of the diazanaphthalenes. The absolute  $pK_1$  value of one compound of the series was determined by recording the pH dependence of the  $^{13}$ C chemical shifts in a dilute solution where the ratio  $f_{RH^+}/f_R$  could be assumed to be unity. The reference value thus obtained enabled us to determine the absolute  $pK_1$  values of the other compounds.

Although the Hammett concept has been the object of criticism (see for example Ref. 5) this procedure for determination of pK values has often been applied with good results (e.g. for primary anilines and indoles). Experimental evidence is available that this concept holds at least for water/sulphuric acid mixtures. Since sulphuric acid is a very strong acid it has the additional advantage that it hardly dilutes the solution when added to set the pH at the desired value.

### Results

In order to determine the relative  $pK_1$  values of the diazanaphthalenes a plot was made of pH vs  $\log I$  (Fig. 1) for all compounds except quinazoline which is not stable in aqueous acid solution. The  $\log I$  value of a compound at a certain pH value was taken as the average of the  $\log I$  values of the separate carbon atoms. Since carbon atoms with a higher  $\Delta\delta$  value provide more accurate  $\log I$  values, only the four carbon atoms with the highest  $\Delta\delta$  values were taken into account.

From Fig. 1 it is seen that the slope of the lines representing the relationship between  $\log I$  and pH is not the same for all compounds. In particular the values for quinoxaline and 1,8-naphthyridine are quite different from the others (0.75 and 1.16, respectively). The slope of the lines for the other compounds is between 0.96 and 1.08.

The difference in  $pK_1$  value between two compounds was determined as the average difference between the log I values—at constant pH value—in the region where the log I values of both compounds were between -1 and +1. The  $pK_1$  value of 1,5-naphthyridine has been determined in dilute solution as a reference value.

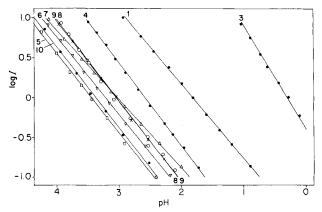


Figure 1. Plot of pH vs Log I for the first nitrogen protonation.

Table 1. Comparison of pK values measured by the <sup>13</sup>C n.m.r. technique with literature values

Compound	p <sub>.</sub> K <sub>1</sub> (n.m.r.)	pK <sub>1</sub> (lit).	Ref.	pK <sub>2</sub> (n.m.r.)	pK <sub>2</sub> (lit.)	Ref.
1	2.20	2.29	11			_
3	0.58	0.56	11	_		_
4	2.93	2.91	11	-1.13	-1.10	13
5	3.81	3.78	10	-0.18	-0.30	13
6	3.57	3.63	10	-1.12		_
7	3.40	3.39	10	-3.10	-2.95	13
8	3.42	3.47	12		_	_
9	3.41	3.48	8	0.37	_	
10	3.74	3.73	а	0.46		_

<sup>&</sup>lt;sup>a</sup> Measured in the same way as in Ref. 10-12

Since the  $pK_1$  values determined with the <sup>13</sup>C n.m.r. technique are in good agreement with the literature values (see Table 1; correlation coefficient c = 0.9990) it seems justified to determine the  $pK_2$  values also with this technique.

For determination of the  $pK_2$  values it is necessary to measure the <sup>13</sup>C chemical shifts of the diprotonated species ( $\delta_3$ ). Since these values may be achieved at very low pH values, dilution effects become important. Therefore the  $pK_2$  value of 1,8-naphthyridine is inaccurate and compounds with lower  $pK_2$  values—i.e. compounds with two nitrogen atoms in one ring—were omitted in this part of our investigation.

The relative  $pK_2$  values were determined from Fig. 2 in the same way as mentioned above for the  $pK_1$  values. The slope of the lines is between 0.53 and 0.80. This indicates that for the latter pH region the proton activity coefficient is smaller than the corresponding coefficient in the pH region of the  $pK_1$  values. The pH dependence of the  $pK_1$  values of 2,7-naphthyridine was also measured in dilute solution, providing the reference  $pK_2$  value.

All pK values are listed in Table 1. From the agreement with the literature values (c = 0.9996) we conclude that the <sup>13</sup>C n.m.r. technique is a good, though time-consuming method for pK measurement.

It is noteworthy that as the separation between the nitrogen atoms increases from 1 carbon atom (1,8-naphthyridine) up to 4 carbon atoms (2,6-naphthyridine) the  $pK_2$  value becomes higher. This may suggest that the  $pK_2$  value is largely determined

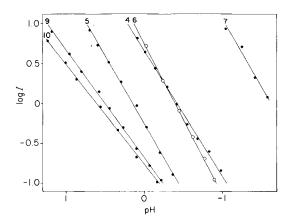


Figure 2. Plot of pH vs log I for the second nitrogen protonation.

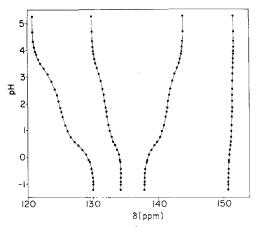


Figure 3. pH dependence of the <sup>13</sup>C chemical shifts of 2,6-naphthyridine.

by the repulsion between the protons. We believe that calculations on the second nitrogen protonation are not significant at the moment, since the experimental structure is not yet known for all molecules (compare  $pK_1$  calculations in Ref. 14). However, we hope to substantiate this suggestion with detailed calculations as soon as the experimental structures of all compounds are known. Single crystal X-ray diffraction measurements are in progress in our laboratory.

# pH DEPENDENCE OF THE <sup>13</sup>C CHEMICAL SHIFTS

### Measurement

In determining the pK values of the diazanaphthalenes we recorded the pH dependence of the  $^{13}$ C chemical shifts for all isomers except quinazoline. As a consequence of the dynamic nature of the protonation equilibrium, protonation does not affect the symmetry of the diazanaphthalenes. Therefore the first and second protonation of the symmetric diazanaphthalenes involve the same, equivalent nitrogen atoms. Thus the change in  $^{13}$ C chemical shifts under the influence of the first nitrogen protonation  $(\Delta \delta_1)$  is similar to that of the second nitrogen protonation  $(\Delta \delta_2)$ ; see for example, 2,6-naphthyridine (Fig. 3). In the asymmetric diazanaphthalenes the first and

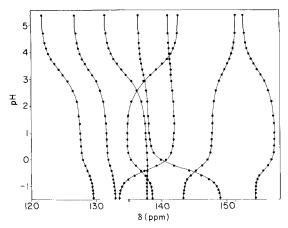


Figure 4. pH dependence of the <sup>13</sup>C chemical shifts of 1,7 naphthyridine.

Table 2. Protonation parameter sets for α- and βnitrogen protonation

C*	$\Delta\delta_{lpha}$	$\Delta\delta_{m{eta}}$
1	b	-2.84
2	-4.67	ь
3	+0.58	-9.61
4	+10.58	+5.08
5	+1.86	+2.36
6	+4.71	+5.71
7	+5.14	+4.27
8	-5.36	+4.02
9	-6.30	+0.33
10	+2.27	+4.18

<sup>&</sup>lt;sup>a</sup> Atom number.

second protonation involve different nitrogen atoms, resulting in different effects on the <sup>13</sup>C chemical shifts; see for example 1,7-naphthyridine (Fig. 4) where the first protonation only affects the  $\beta$ -nitrogen atom and the second protonation the  $\alpha$ -nitrogen atom.

The <sup>13</sup>C chemical shifts of the neutral molecules  $(\delta_1)$ , monoprotonated molecules  $(\delta_2)$ , diprotonated molecules  $(\delta_3)$  and the changes in <sup>13</sup>C chemical shifts  $(\Delta \delta_1 \text{ and } \Delta \delta_2)$  are listed in Tables 3 and 4. For the molecules which have both nitrogen atoms in one ring (1, 3 and 8) the p $K_2$  value is so low that the  $\delta_3$  values could not be measured by our method due to excessive dilution.

Table 3. pH dependence of the <sup>13</sup>C chemical shifts in the symmetric diazanaphthalenes

C*	$\delta_1^{b}$	82 <sup>b</sup>	$\delta_3$	$\Delta {\delta_1}^{\mathbf{b}}$	$\Delta \delta_2$	Δδ(exp) <sup>c</sup>
<b>3</b> -2/3	145.19	143.22		-1.97	_	-2.04
<b>3</b> -5/8	128.42	126.68	_	-1.74	_	-1.74
<b>3</b> -6/7	131.58	136.62	_	+5.04	_	+4.93
<b>3</b> -9/10	141.17	138.73	_	-2.44	_	-2.02
4-2/6	151.31	151.39	151.55	+0.08	+0.16	+0.02
4-3/7	125.72	128.68	131.64	+2.96	+2.96	+2.86
<b>4</b> -4/8	136.99	139.60	142.16	+2.61	+2.56	+2.61
4-9/10	141.78	139.60	135.30	-2.18	-4.30	-2.02
<b>7</b> -2/7	153.85	154.00	153.60	+0.15	-0.40	+0.23
<b>7</b> -3/6	123.54	126.06	128.83	+2.52	+2.77	+2.64
<b>7</b> -4/5	139.27	145.50	153.40	+6.23	+7.90	+6.23
<b>7</b> -9	153.37	147.25	139.10	-6.12	-8.15	-6.30
<b>7</b> -10	123.15	125.60	127.22	+2.45	+1.62	+2.27
8-1/4	152.01	153.23		+1.22		+1.12
<b>8</b> -5/8	127.23	130.52		+3.29	_	+3.19
<b>8</b> -6/7	134.67	139.16	_	+4.49	_	+4.49
<b>8</b> -9/10	126.73	128.99		+2.26		+2.26
<b>9</b> -1/5	151.53	151.24	150.68	-0.29	-0.56	~0.24
<b>9-</b> 3/ <b>7</b>	143.87	141.20	137.90	-2.67	-3.30	-2.67
<b>9</b> -4/8	120.73	125.23	130.07	+4.50	+4.84	+4.55
<b>9</b> -9/10	129.82	132.07	134.24	+2.25	+2.17	+2.26
<b>10</b> -1/8	152.61	153.14	153.49	+0.53	+0.35	+0.59
<b>10</b> -3/6	146.24	144.29	141.50	-1.95	<b>-2.7</b> 9	-1.95
10-4/5	120.58	124.24	127.83	+3.66	+3.59	+3.72
<b>10</b> -9	123.27	123.86	123.99	+0.59	+0.13	+0.33
<b>10</b> -10	138.57	143.01	146.49	+4.44	+3.48	+4.18

<sup>\*</sup> Compound and carbon atom number.

<sup>&</sup>lt;sup>b</sup> Nitrogen atom.

<sup>&</sup>lt;sup>b</sup> Results concerning the first protonation are slightly different from the values given in Ref. 1 because of different experimental conditions (see Experimental).

<sup>&</sup>lt;sup>c</sup> Expected values from protonation parameters.

Table 4. pH dependence of the <sup>13</sup>C chemical shifts in the asymmetric diazanaphthalenes

C.	$\delta_1^{\mathrm{b}}$	$\delta_2^{\ b}$	$\delta_3$	$\Delta \delta_1^{\ b}$	Δδ <sub>1</sub> <sup>c</sup> (exp)	$\Delta\delta_2$	$\Delta \delta_2^{c}(exp)$
1-3	144.93	141.14		-3.78	−5.16 <sup>e</sup>	_	
1-4	125.87	135.46	_	+9.59	+7.50	_	
1-5	127.36	128.78	-	+1.42	+2.14		_
1-6	132.75	138.83	_	+6.08	+5.27	_	_
1-7	132.68	137.85	_	+5.17	+4.65	_	
1-8	127.75	128.40	_	+0.75	-0.07	_	_
1-9	149.33	148.73		-0.60	-2.56	_	
<b>1</b> -10	126.86	132.96		+6.10	+3.35		_
<b>5</b> -2	155.66	161.14	156.70	+5.48	+5.71	-4.44	-4.67
<b>5</b> -3 <sup>d</sup>	124.16	127.30	129.10	+3.14	+4.27	+1.80	+0.58
5-4	137.80	141.60	152.60	+3.80	+4.02	+11.00	+10.58
<b>5</b> -5	152.95	150.24	152.25	-2.71	-2.84	+2.01	+1.86
<b>5</b> -7	145.91	138.10	141.65	-7.81	-9.61	+3.55	+5.14
<b>5</b> -8 <sup>d</sup>	121.64	126.39	121.70	+4.75	+5.08	-4.69	-5.36
5-9	148.67	151.72	144.55	+3.05	+4.18	-7.17	-6.31
<b>5</b> -10	123.48	125.01	125.78	+1.53	+0.33	+0.77	+2.27
<b>6</b> -2	152.53	157.33	154.43	+4.80	+4.02	-2.90	-4.67
<b>6</b> -3	126.74	131.51	132.95	+4.77	+5.71	+1.44	+0.58
6-4	136.50	138.05	149.15	+1.55	+2.36	+11.10	+10.58
<b>6</b> -5	121.70	127.50	129.60	+5.80	+5.08	+2.10	+1.86
<b>6</b> -6	142.63	134.67	138.70	-7.96	-9.61	+4.03	+4.71
<b>6</b> -8	151.35	147.77	143.27	-3.58	-2.84	-4.50	-5.36
<b>6</b> -9	141.03	141.92	133.38	+0.89	-0.33	-8.54	-6.30
<b>6</b> -10	131.37	137.51	137.78	+6.14	+4.18	+0.27	+2.28

a-c See Table 3.

The first nitrogen protonation of the symmetric diazanaphthalenes provides us with two protonation parameter sets, one for  $\alpha$ - and one for  $\beta$ -nitrogen protonation (Table 2.).

These protonation parameters give a very good description of the  $\Delta\delta_1$  values (Tables 3 and 4; c=0.986). The  $\Delta\delta_2$  values could also be described quite well by these protonation parameters (Table 3 and 4; c=0.978). This is in contrast with the results which Pugmire and Grant<sup>15</sup> have reported for the diazabenzenes. However, we have to realize that the diazabenzenes contain two nitrogen atoms in one ring, whereas

we have not been able to measure the second protonation of the corresponding diazanaphthalenes.

### **EXPERIMENTAL**

The proton noise-decoupled <sup>13</sup>C n.m.r. spectra were recorded on a Varian XL-100 spectrometer (25.2 MHz for <sup>13</sup>C and 15.4 MHz for <sup>2</sup>H lock). The data acquisition of the free induction delays (pulse width 22  $\mu$ s; acquisition time 0.8 s; pulse delay 0.5 s) and Fourier transformation were performed with a Varian 620/L data machine (16 K). With a spectral width of 5120 Hz and 4096 memory points the resolution is 0.05 ppm. Using water as a solvent and concentration of  $(150\pm1)$  mg ml<sup>-1</sup>, 500 transients were required to obtain an acceptable signal to noise ratio. The measurements for the reference pK values were performed on solutions with concentrations of  $(15\pm0.5)$  mg ml<sup>-1</sup>. In this case 30 000 transients were required for each <sup>13</sup>C n.m.r. spectrum.

In the tube (o.d. 10 mm) containing the aqueous solution, a capillary was centred with the aid of three Teflon rings. The capillary contained deuterated acetone to provide a signal for the deuterium field-frequency lock and TMS as an internal standard.

The pH was decreased progressively with sulphuric acid (98%, Merck) and measured with a Knick pH-meter at 20 °C (probe temperature  $20\pm2$  °C) with an Ingold combined electrode (type 405M5). This electrode was calibrated with Elektrofact buffer powder at pH=4 and pH=7. Compounds 1, 3 and 8 are commercial products (Aldrich). The other compounds were synthesized according to Refs. 16–20.

### Acknowledgement

We wish to thank Professor Dr D. Feil for critically reading the manuscript.

The investigations were supported in part by the Netherlands Foundation for Chemical Research (SON) with financial aid from the Netherlands Organization for the Advancement of Pure Research (ZWO).

# REFERENCES

Received 7 July 1976; accepted 13 October 1976

© Heyden & Son Ltd, 1977

<sup>&</sup>lt;sup>d</sup> Because of their  $\Delta\delta_2$ -values the assignment of C-3 and C-8 has been reversed compared with Ref. 1.

<sup>•</sup>  $\Delta\delta_1(\exp)$  for cinnoline: 0.44  $\Delta\delta_{\alpha}$  + 0.56 $\Delta\delta_{\beta}$  (see Ref. 1).

P. van de Weijer, H. Thijsse and D. van der Meer, Org. Magn. Reson. 8, 187 (1976).

J. S. Cohen, R. I. Schrager, M. McNeel and A. N. Schechter, Nature (London) 228, 642 (1970).

<sup>3.</sup> E. Breitmaier and K.-H. Spohn, Tetrahedron 29, 1145 (1973).

L. P. Hammett and A. J. Deyrup, J. Am. Chem. Soc. 54, 2721 (1932).

M. M. Kreevoy and E. H. Baughman, J. Am. Chem. Soc. 95, 8178 (1974).

M. J. Jorgenson and D. R. Hartten, J. Am. Chem. Soc. 85, 878 (1963).

<sup>7.</sup> R. L. Hinman and J. Lang, *J. Am. Chem. Soc.* **86**, 3796 (1964).

<sup>8.</sup> J. F. Bunnett and F. P. Olsen, *Can. J. Chem.* 44, 1899 (1966).

<sup>9.</sup> A. Albert, D. J. Brown and H. C. S. Wood, *J. Chem. Soc.* 3832 (1954).

<sup>10.</sup> A. Albert, J. Chem. Soc. 1790 (1960).

<sup>11.</sup> A. Albert and J. N. Philips, J. Chem. Soc. 1294 (1956).

A. Albert, W. L. F. Armarego and E. Spinner, J. Chem. Soc. 2689 (1961).

E. Laviron and L. Roullier, C.R. Acad. Sci. Ser. C. 274, 1489 (1972).

P. van de Weijer, D. van der Meer and J. L. Koster, Theor. Chim. Acta 38, 223 (1975).

R. J. Pugmire and D. M. Grant, J. Am. Chem. Soc. 90, 697 (1968).

T. J. Kress and W. W. Pandler, Chem. Commun. 1, 3 (1967), 1,6-naphthyridine.
 H. Rapoport and A. Batcho, J. Org. Chem. 28, 1753 (1963),

<sup>1,7-</sup>naphthyridine.

18. T. J. Kress and W. W. Paudler, *J. Org. Chem.* 32, 832 (1967),

<sup>1,8-</sup>naphthyridine.
19. A. Taurins and R. Tanli, Can. J. Chem. 52, 843 (1974),

<sup>2,6-</sup>naphthyridine.

W. W. Paudler and S. J. Cornich, J. Heterocycl. Chem. 7, 419 (1970), 2,7,-naphthyridine.